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# Leveraging 3D *In Vitro* Models for Oncology Drug Discovery

**Rajendra Kumari, PhD**

**Global Head of Scientific Communications**



# Speaker

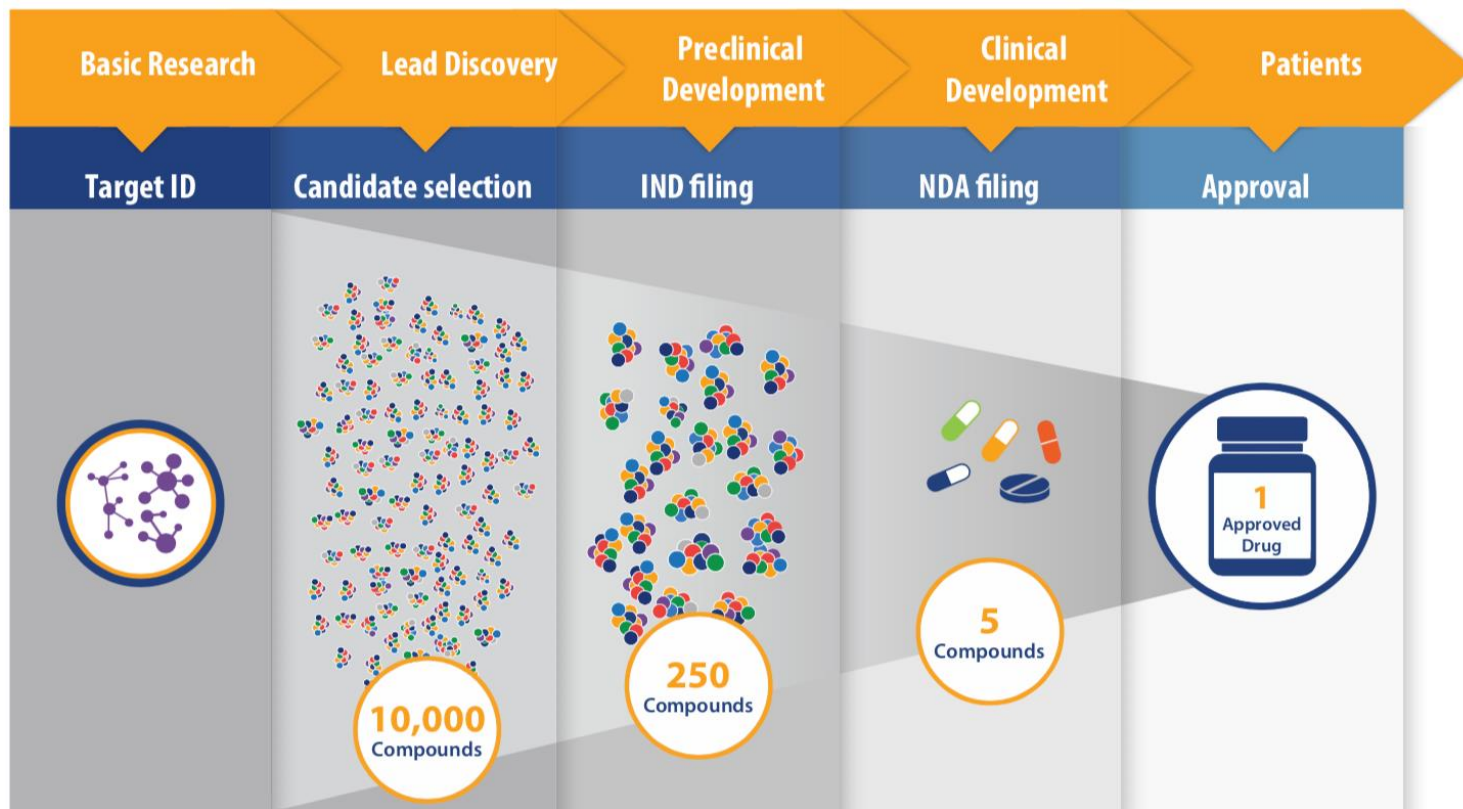
## Rajendra Kumari, PhD

- Received her PhD from the University of Leicester in Molecular Pharmacology, followed by postdoctoral fellowships in the Division of Preclinical Oncology, University of Nottingham, and lectureship in the School of Clinical Sciences
- Co-founder of PRECOS Ltd in 2010, a spin-out company providing preclinical oncology services for *in vitro* and *in vivo* research, as well as imaging, taking the role of Chief Operations Officer
- PRECOS merged with CrownBio in 2013, where Rajendra served as General Manager and Chief Scientific Officer of CrownBio's European facility in the UK
- Currently is the Global Head of Scientific Communication
- Rajendra is a member of the American Association of Cancer Research and the British Association of Cancer Research, and has authored over 40 abstracts and publications



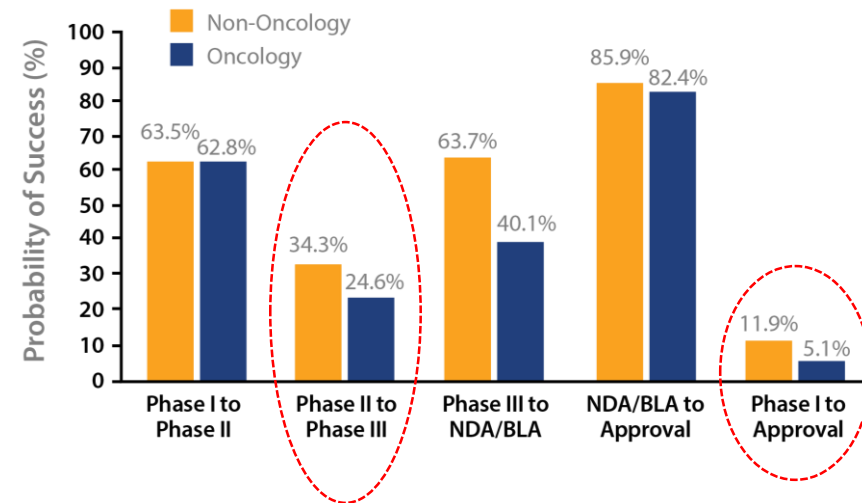
- Bridging the gap between the lab and the clinic
- Key advantages of using 3D *in vitro* systems
- When to use 3D *in vitro* platforms, including new technology like organoids
- How to integrate organoids into your drug discovery program and when to look for alternatives
- Predicting *in vivo* responses to move forward with

# Current Linear Drug Discovery Process: Inefficient, Costly and Risky



## Cancer drug development

- Most candidates fail
- Attrition rates particularly high



*Nat Rev Drug Discov, 2004, 3:711*

# Preclinical Tumor Models for Oncology Drug Discovery

## Historical models



2D cell lines



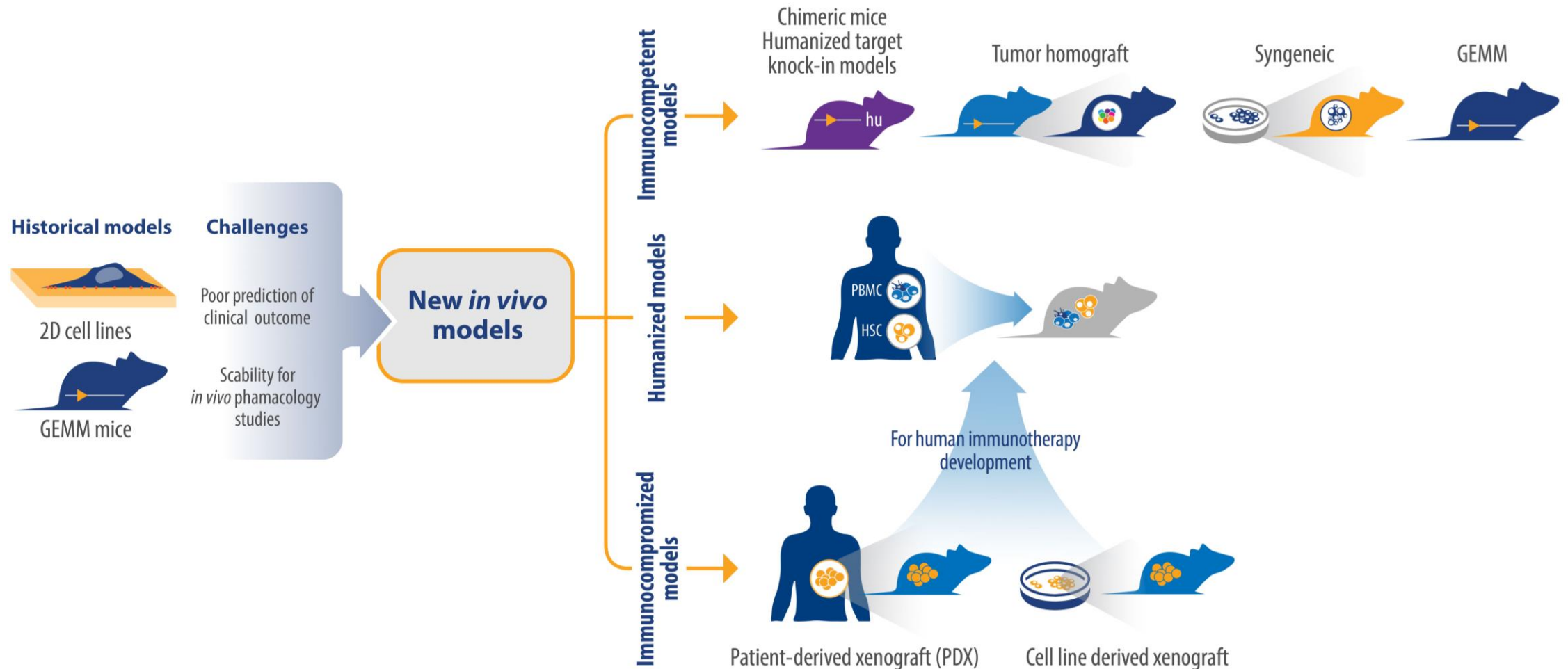
GEMM mice

## Challenges

Poor prediction of  
clinical outcome

Scability for  
*in vivo* phamacology  
studies

# Preclinical Tumor Models for Oncology Drug Discovery



# Patient-Derived Models Bridge the Gap Between Lab and Clinic

- Preserve original tumor histo- and molecular pathology
- Recapitulate intra- and inter-patient heterogeneity, and patient diversity
- Pharmacologically predictive

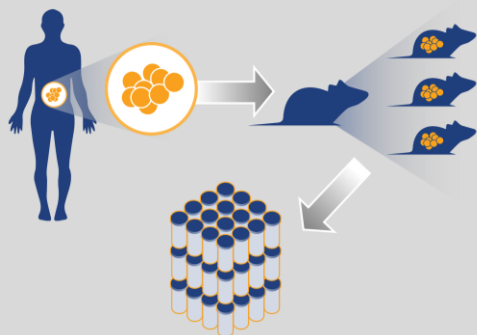




# The Value of Patient-Derived Model Biobanks

*In Vivo*

## PDX development and biobanking



## Large, diverse, and well characterized biobank

From **US**, **European**, and **Asian** populations



30 different **cancer types**

## PDX Mouse Clinical Trial (MCT)

**RCT-MCT: "1" or "n+n"**  
Multiple Indications and Targets



**Data analysis**  
(Paired survival analysis)

## HuPrime®

~2500 PDX  
established and  
~1500 RNASeq'd



## HuBase™

Online searchable PDX  
database to access  
genomic annotation



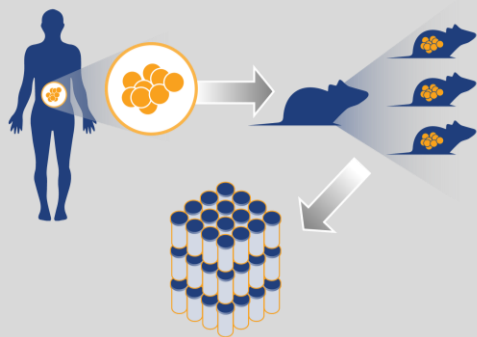
Register at: <https://www.crownbio.com/hb-registration>





# The Value of Patient-Derived Model Biobanks

## In Vivo



**Large, diverse, and well characterized biobank**

From **US**, **European**, and **Asian** populations



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## PDX Mouse Clinical Trial (MCT)

## RCT-MCT: "1" or "n+n"

### Multiple Indications and Targets



## Data analysis

(Paired survival analysis)

HuPrime®

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HuBase™

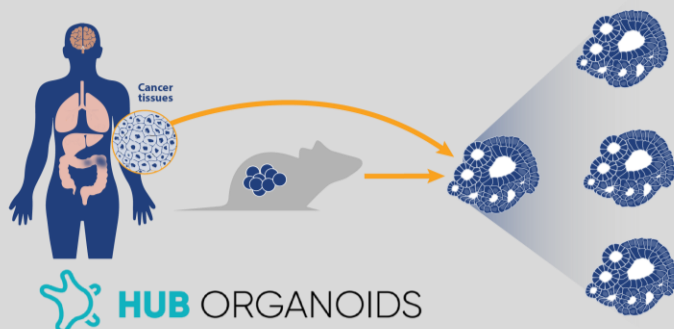
Online searchable PD database to access genomic annotation



Register at: <https://www.crownbio.com/hb-registration>

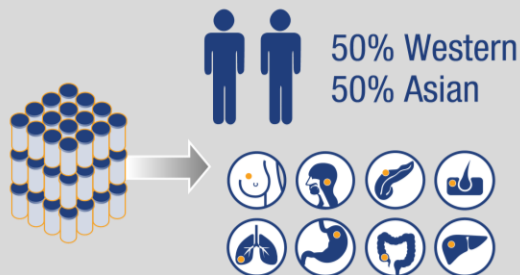
## In Vitro 3D

## Organoid development and biobanking



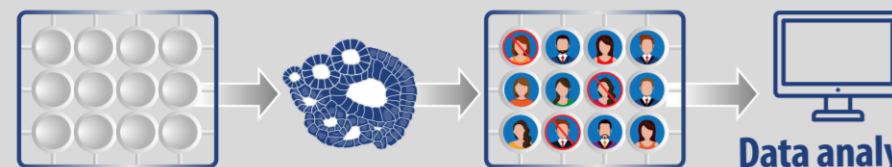
## Large, diverse, and well characterized biobank

From Western and Asian populations



15 different cancer types

## Clinical Trial in a Dish (CTiD)



Register at: <https://www.crownbio.com/ob-registration>

## Integration into the Drug Development Process



**How can you leverage patient-derived *in vitro* models?**



**How to integrate patient-derived 3D models into drug development?**



**What advantage will they offer?**



**When do you use these models?**



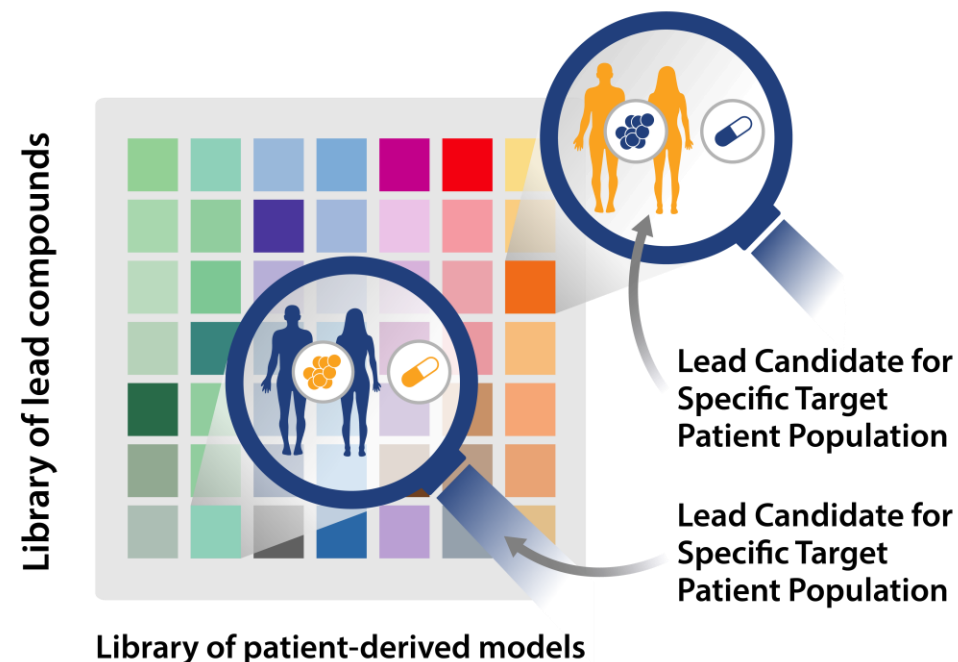
**How can they be used for immuno-oncology applications?**



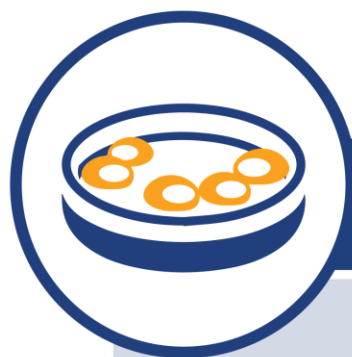
**How do I access them?**

# An Improved Workflow in Oncology Drug Discovery is Needed

- Drug discovery requires selection on “**two variables**”
  - **A lead compound** by screening large numbers of drug candidates
  - **A target patient population** (indication) by screening a diverse portfolio of disease models
- Accelerate and simplify clinical development by screening **lead/model libraries** simultaneously via a **matrix high-throughput screen** (HTS)
- Identify potential **CDx** or **biomarkers** of response
- **Mouse clinical trials** have inherent limitations in handling this type of screening, e.g. time, cost



# *In Vitro/Ex Vivo* Preclinical Tumor Models



## 2D systems

- Primary cell cultures
- Standard cell lines
- Engineered lines
- Co-incubation



## 3D systems

- Patient-derived organoids
- *Ex vivo* cell cultures
- Standard cell lines
- Engineered lines
- Co-cultures



## *Ex vivo* tissue

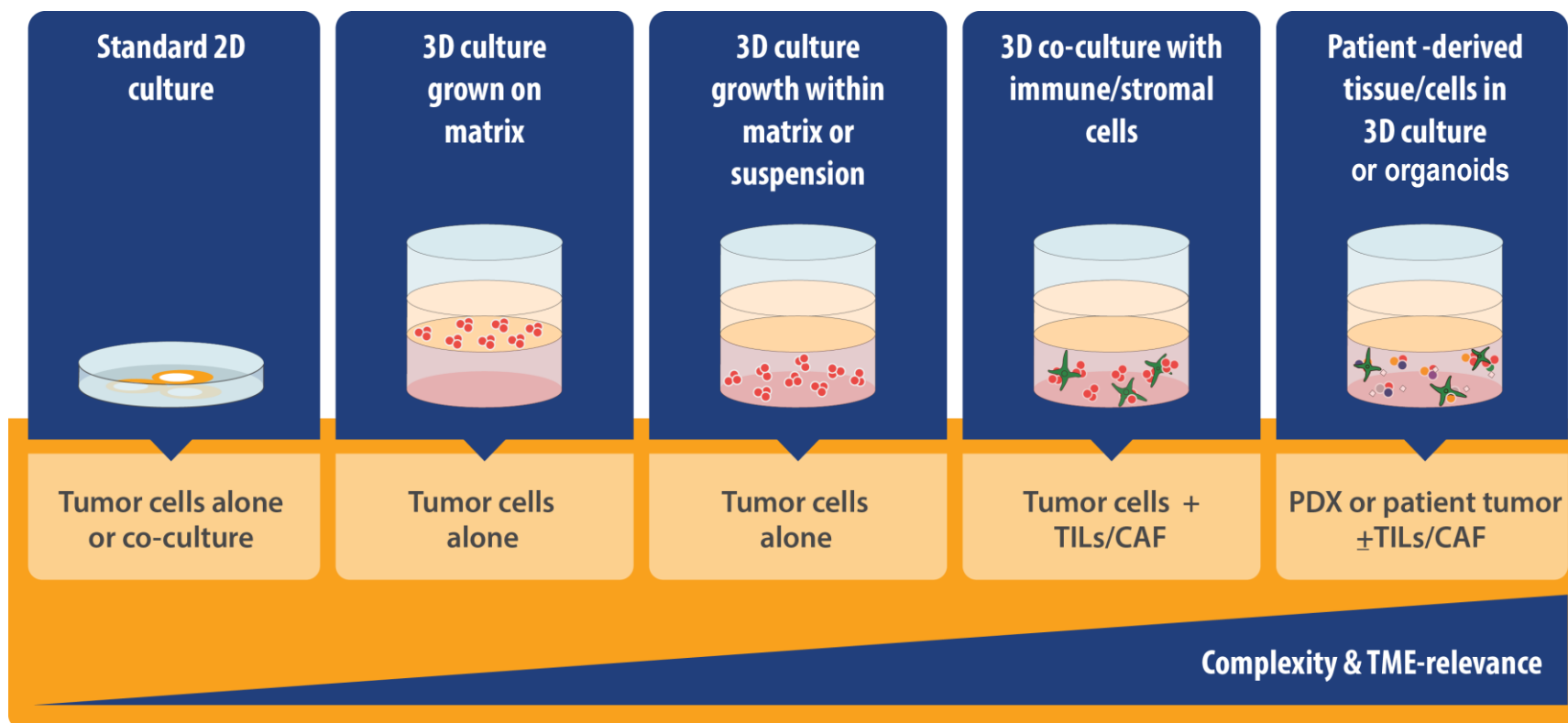
- Patient-derived
- Animal-derived

Navigate Between 2D and 3D Systems

# **SELECTING IN VITRO SYSTEMS FOR ONCOLOGY DRUG DISCOVERY**

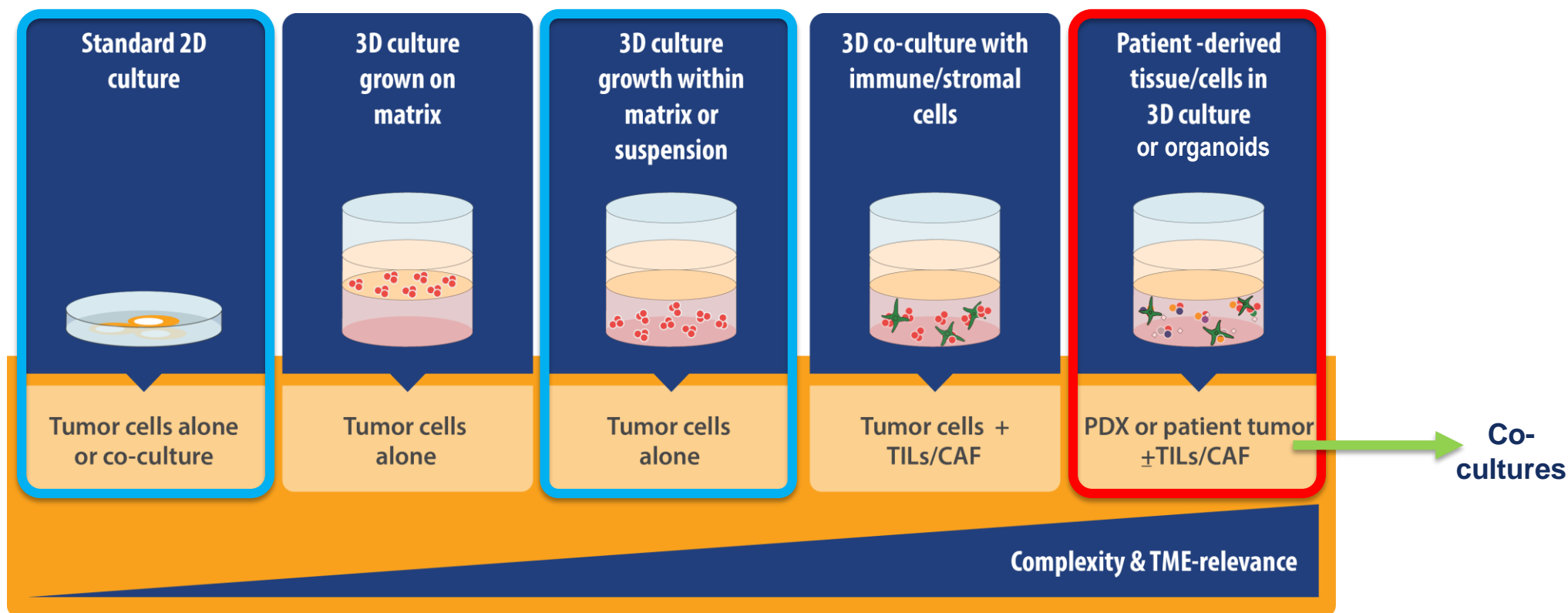
# Tumor Microenvironment (TME) Relevance

- 3D modelling increases patient relevance and potentially impacts complexity and throughput



# Tumor Microenvironment (TME) Relevance

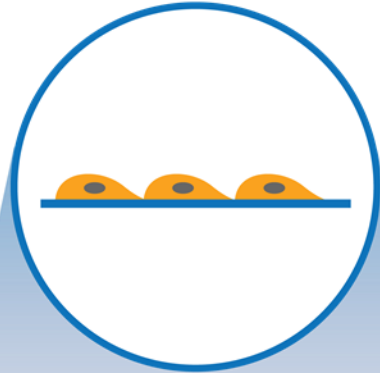
- 3D modelling increases patient relevance and potentially impacts complexity and throughput





# Key Features

## 2D culture



Derived from  
cell line monocultures

Single cell population

Monolayer growth

Long term culture

## 3D Spheroid



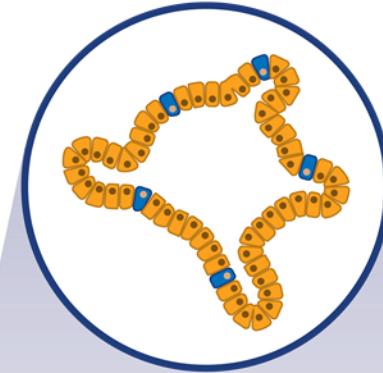
Derived from cell line  
cultures or tissue

Represent single/  
partial tissue components

Transiently resemble  
cell organization

Difficult to maintain long term

## Organoid



Derived from stem cells

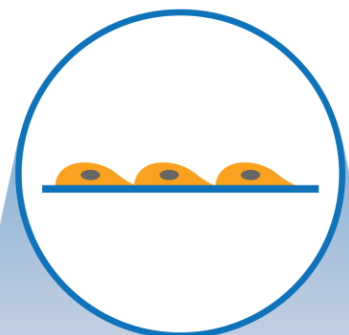
Multiple cell lineages

Recapitulate organ  
physiological parameters

Long term culture

# Why Use 2D Cell-Based Models?

## 2D culture



Derived from  
cell line monocultures

Single cell population

Monolayer growth

Long term culture

## Advantages

- Robust & reproducible, with high performance & low cost
- Simple and long-term culturing, amenable to engineering
- Large collection of well published models available
- Commercial tests widely available

## Limitations

- Poor translatability *in vivo*, as they do not mimic the natural tumor or tissue structure
- Limited heterogeneity and diversity

## Applications

- Investigate target engagement, oncogenic pathways, MoA
- HTS – lead identification
- Workhorse models for lead optimization
- Non-translational studies

When to Choose Organoids

# **SELECTING A RELEVANT 3D SYSTEM**

# Difference Between Spheroids and Organoids

## 3D Spheroid



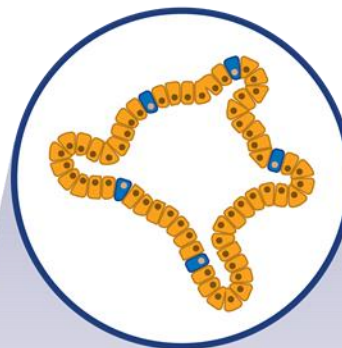
Derived from cell line  
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## Organoid

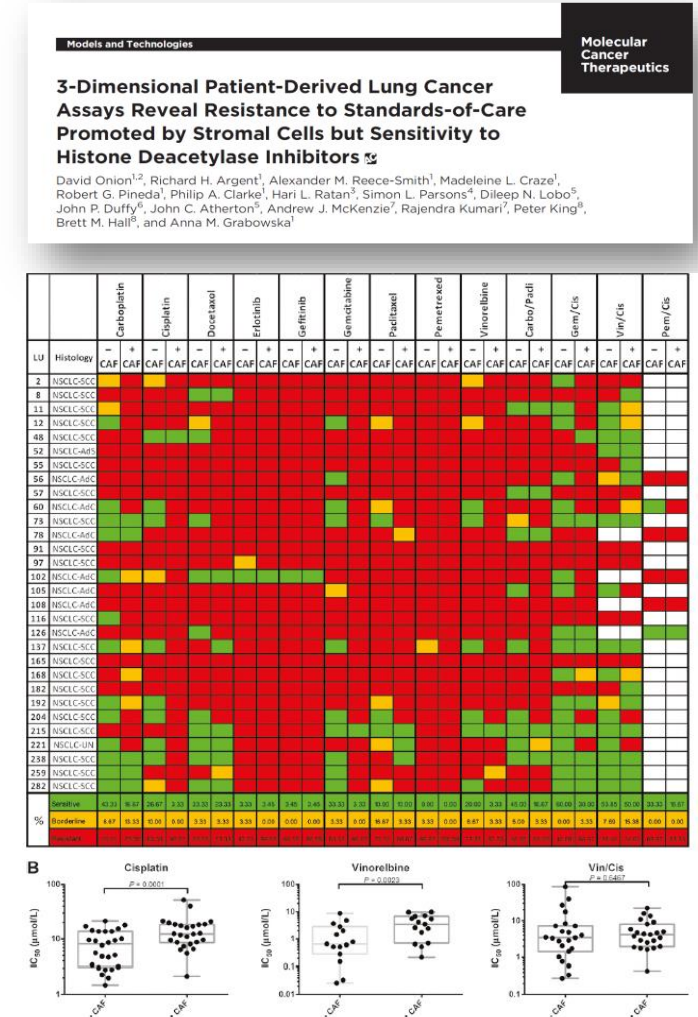


Derived from stem cells

Multiple cell lineages

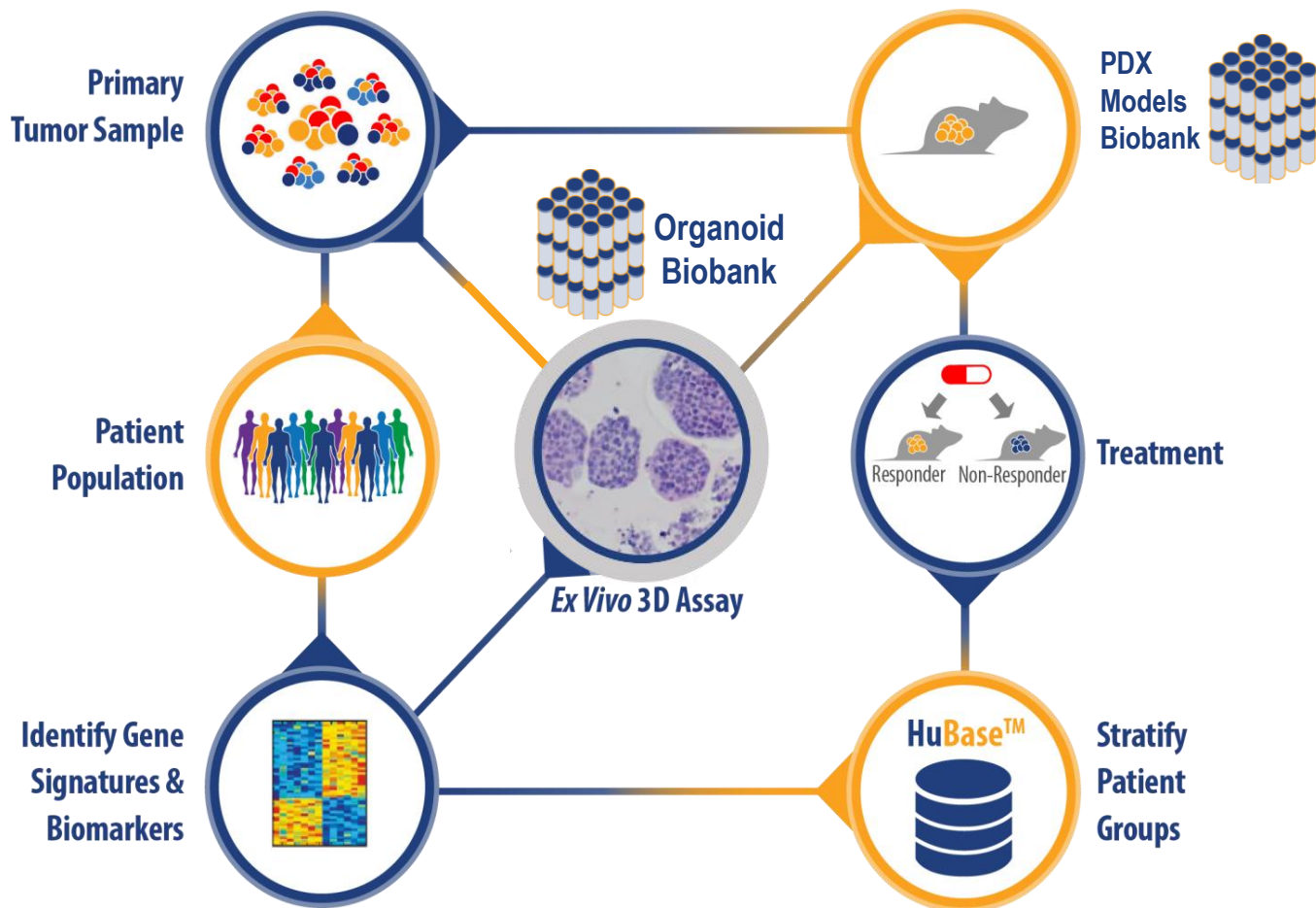
Recapitulate organ  
physiological parameters

Long term culture



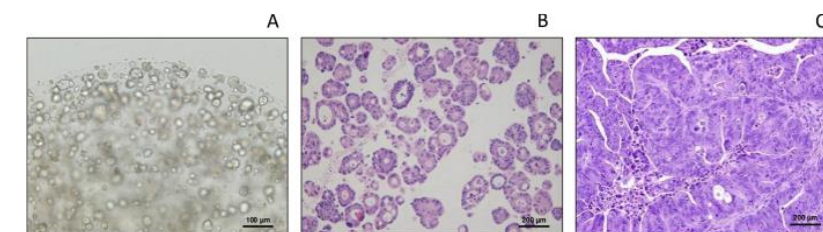


# 3D Screen Using *In Vitro* Organoids

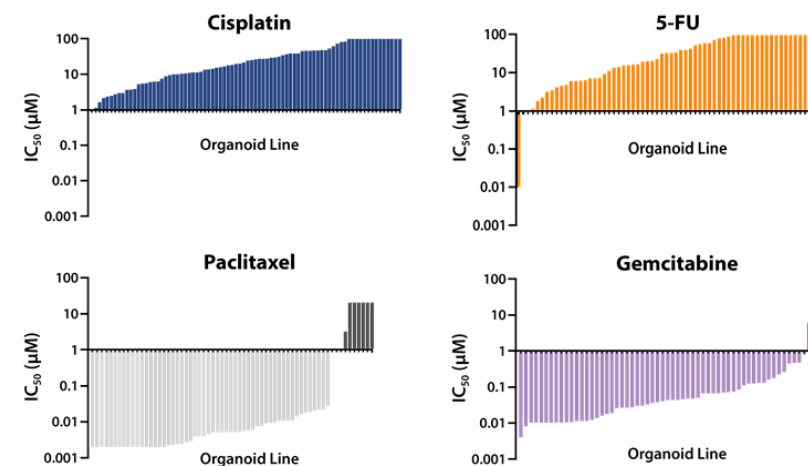


## Creating Matched In vivo/In vitro Patient-Derived Model Pairs of PDX and PDX-Derived Organoids for Cancer Pharmacology Research

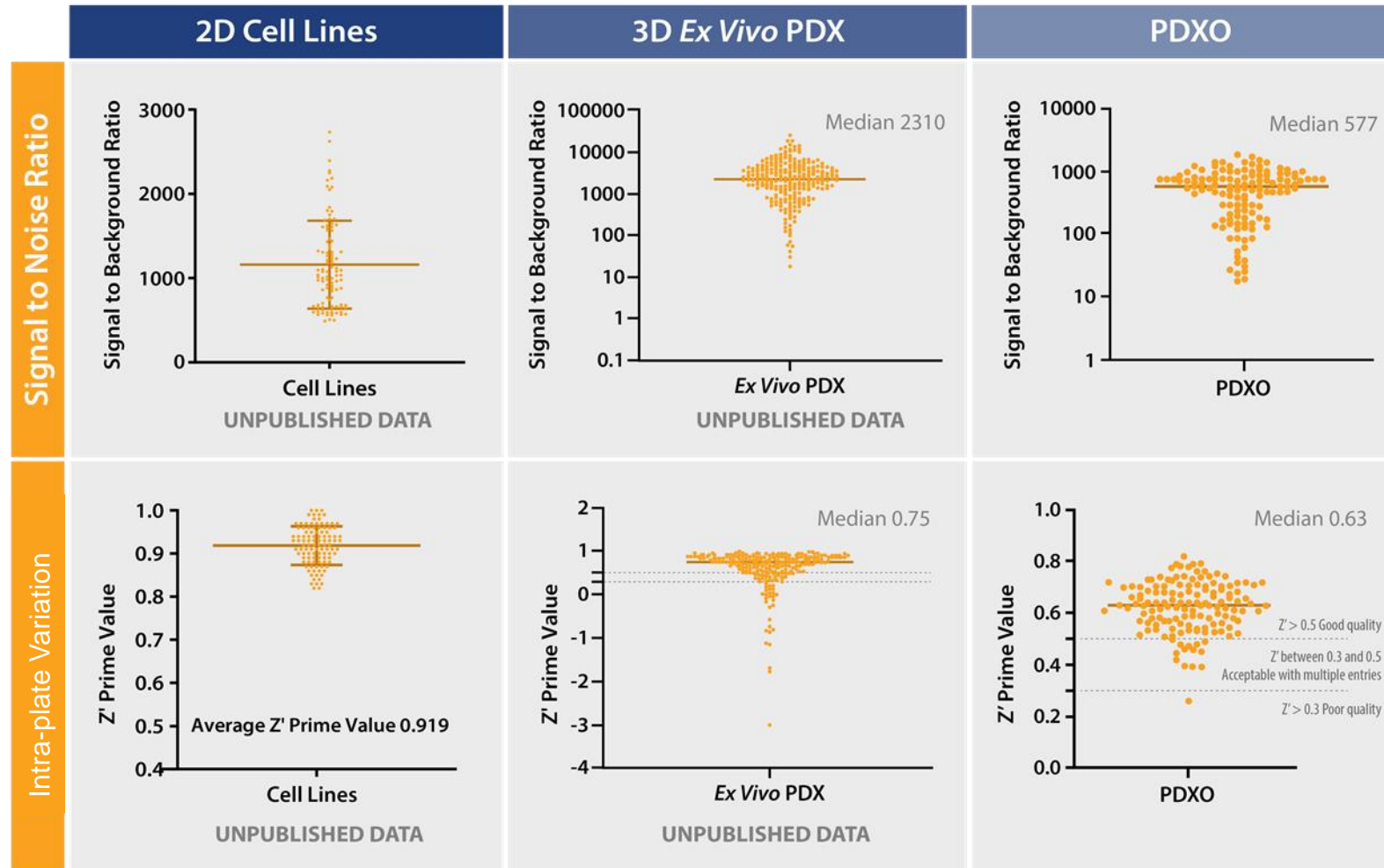
Xiaoxi Xu<sup>1</sup>, Limei Shang<sup>1</sup>, Shuzong Wang<sup>2</sup>, Jun Zhou<sup>2</sup>, Xuesong Ouyang<sup>2</sup>, Meiling Zheng<sup>1</sup>, Binchen Mao<sup>2</sup>, Likun Zhang<sup>2</sup>, Xiaobo Chen<sup>1</sup>, Jingjing Wang<sup>2</sup>, Jing Chen<sup>3</sup>, Wubin Qian<sup>2</sup>, Sheng Guo<sup>2</sup>, Yujun Huang<sup>2</sup>, Qi-Xiang Li<sup>3</sup>



Xu et al 2020 JOVE in press



# Assay Validation Comparison

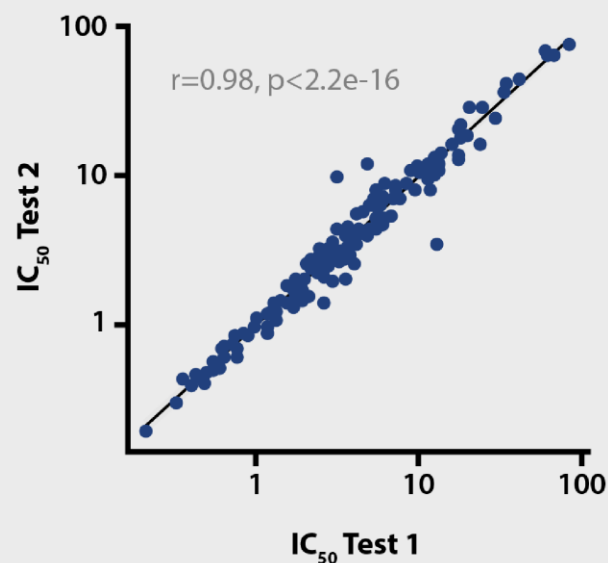




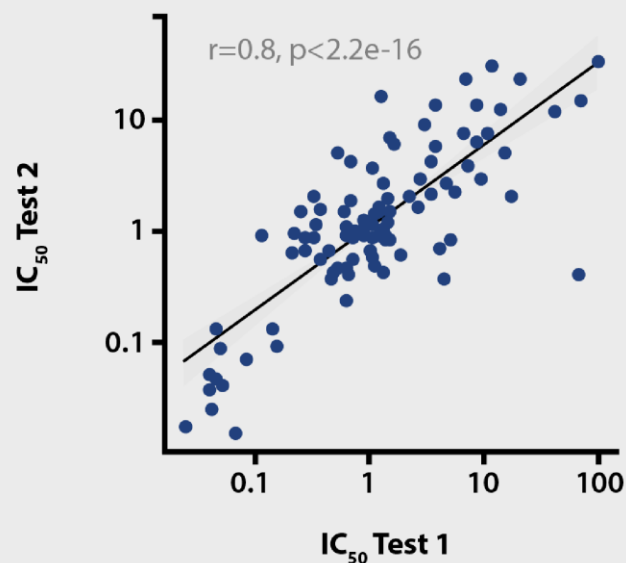
# Inter-plate Variation Comparison

Inter-plate Variation ( $IC_{50}$ )

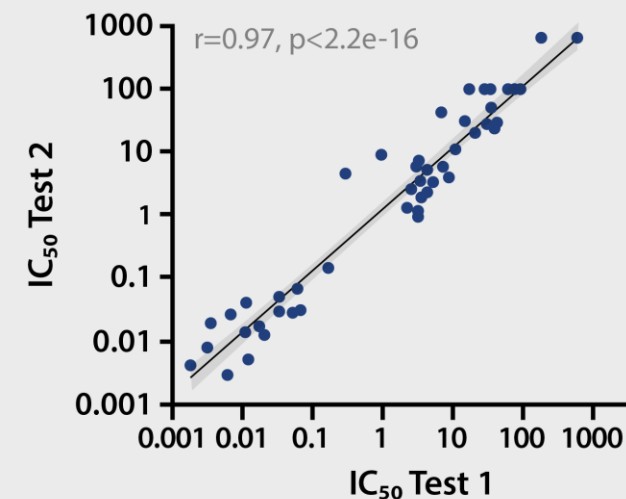
2D Cell Lines



3D *Ex Vivo* PDX



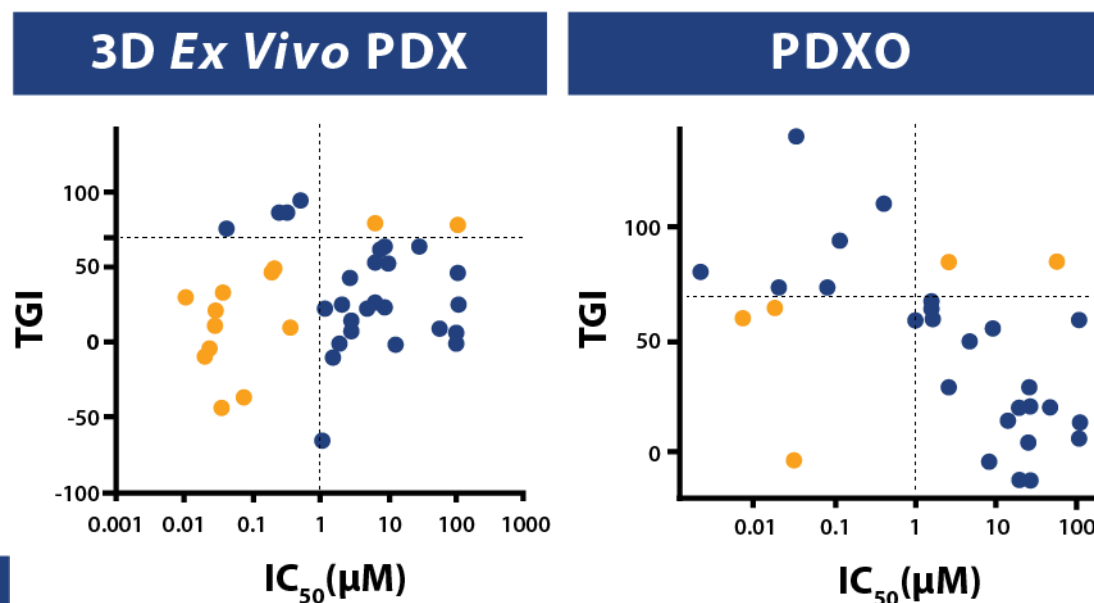
PDXO



UNPUBLISHED DATA

# Correlation with *In Vivo* Response

- PDXO have a higher prediction power of corresponding *in vivo* PDX response compared to 3D *ex vivo* PDX assay



Overall prediction 68%  
Fisher's exact test p value = 0.17

TGI	IC <sub>50</sub>	
	Sensitive (IC <sub>50</sub> < 1 μM)	Resistant (IC <sub>50</sub> ≥ 1 μM)
Sensitive (TGI ≥ 70%)	4 (Positive prediction 27%)	2
Resistant (TGI < 70%)	11	23 (Negative prediction 92%)

Overall prediction 86%  
Fisher's exact test p value = 0.001135

TGI	IC <sub>50</sub>	
	Sensitive (IC <sub>50</sub> ≥ 1 μM)	Resistant (IC <sub>50</sub> ≥ 1 μM)
Sensitive (TGI ≥ 70%)	6 (Positive prediction 75%)	2
Resistant (TGI < 70%)	2	20 (Negative prediction 91%)

UNPUBLISHED DATA

# Why Use 3D Cell-Based Models?

## 3D Spheroid



Derived from cell line cultures or tissue

Represent single/partial tissue components

Transiently resemble cell organization

Difficult to maintain long term

### Advantages

- Suitable for wide range of cells/tissue & co-culture conditions
- More closely mimic tissue architecture
- Greater physiological relevance
- Enhanced translatability, compared to 2D models

### Limitations

- Unstable in long-term culture, due to cellular senescence
- Lower reproducibility and translatability *in vivo*
- Limited tissue availability can challenge scalability
- Automation achievable but lower throughput than 2D

### Applications

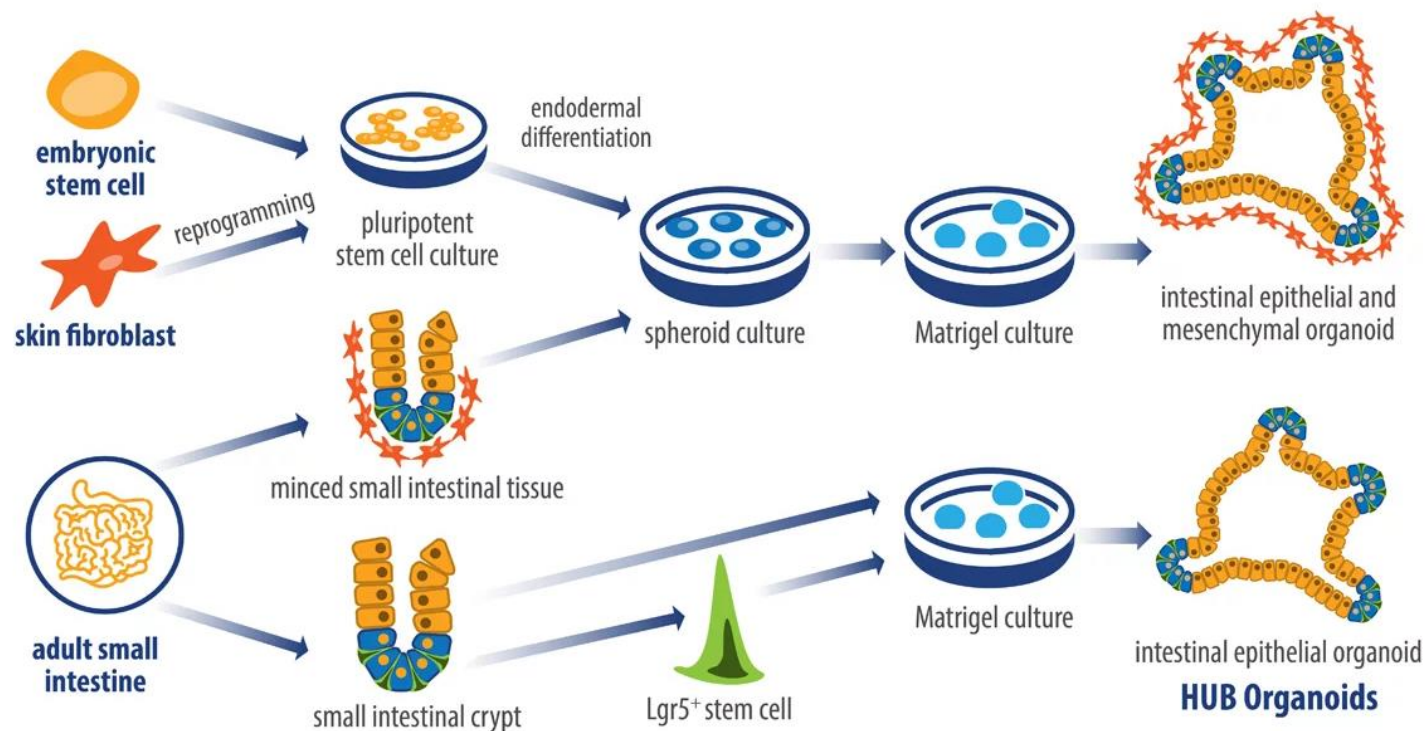
- Investigate efficacy, target engagement, oncogenic pathways, MoA in relevant TME
- CSCs, hypoxia and drug penetrance
- Clinical hypotheses testing and screening in primary tissue

Quick Tips

**WHICH ORGANOID SYSTEM?**

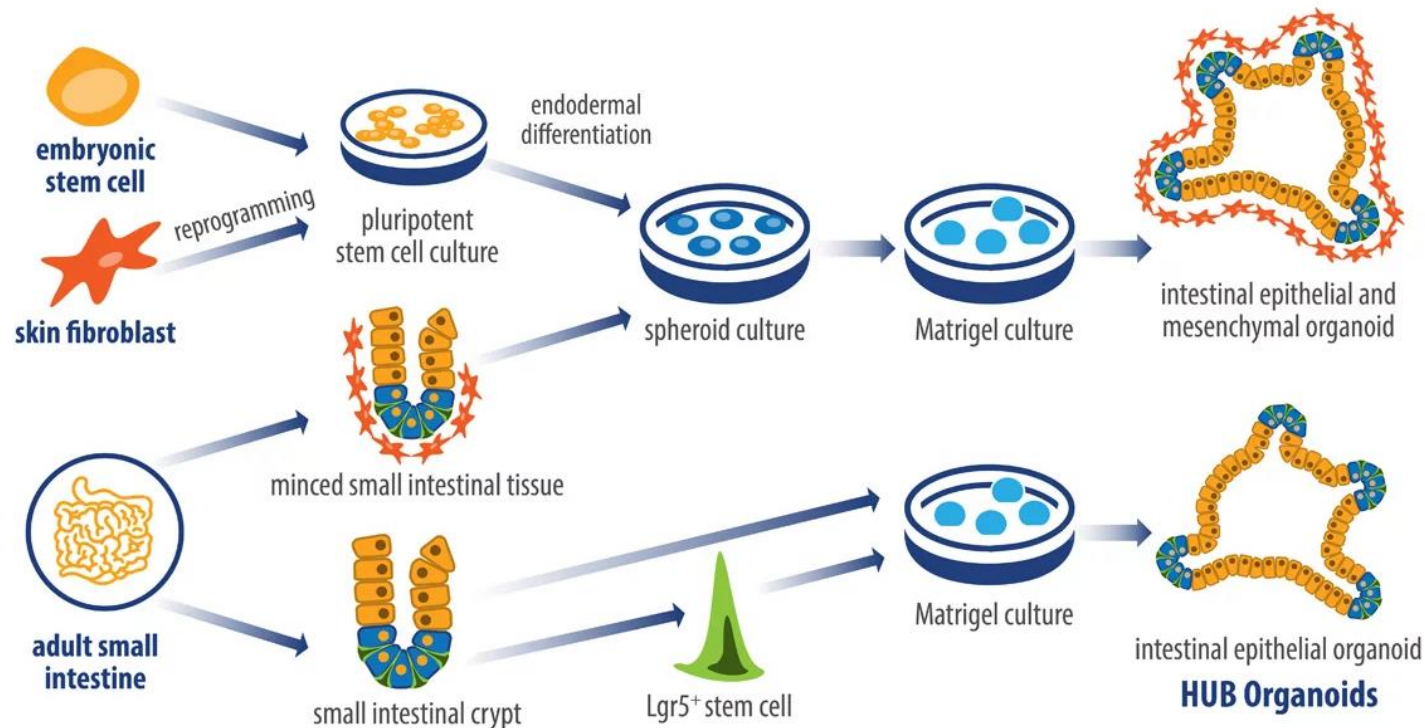
# The advantage of HUB Organoid Technology

- Pure epithelial compartment; absence of mesenchymal cellular niche
- More robust and genomically stable; faster establishment
- The only **cancer patient-relevant** technology

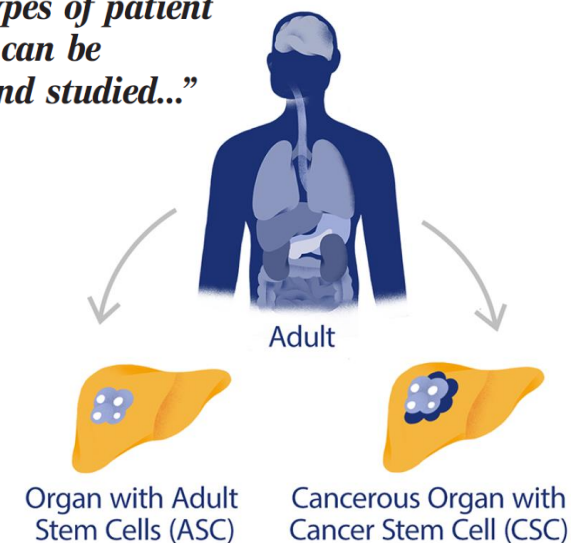


# The advantage of HUB Organoid Technology

- Pure epithelial compartment; absence of mesenchymal cellular niche
- More robust and genomically stable; faster establishment
- The only **cancer patient-relevant** technology



*“...PDO biobanks greatly expand the types of patient samples that can be propagated and studied...”*





# Organoids Predict Clinical Response

- Translatable models with demonstrated predictive power of clinical response

Vlachogiannis *et al.*, *Science* **359**, 920–926 (2018)

RESEARCH | REPORTS

## ORGANOIDS

### Patient-derived organoids model treatment response of metastatic gastrointestinal cancers

Georgios Vlachogiannis,<sup>1</sup> Somaieh Hedayat,<sup>1</sup> Alexandra Vatsiou,<sup>2</sup> Yann Jamin,<sup>3</sup> Javier Fernández-Mateos,<sup>1,2</sup> Khurum Khan,<sup>1,4</sup> Andrea Lampis,<sup>1</sup> Katherine Eason,<sup>1</sup> Ian Huntingford,<sup>1</sup> Rosemary Burke,<sup>5</sup> Mihaila Rata,<sup>3</sup> Dong-Mu Koh,<sup>3,6</sup> Ning Tian,<sup>3,6</sup> David Collins,<sup>3</sup> Sanna Hulkki-Wilson,<sup>1</sup> Chao Sing Yu Moorcraft,<sup>4</sup> Ian Chau,<sup>4</sup> Sheela Rao,<sup>4</sup> Maria Bali,<sup>3,6</sup> Mahnaz Darvish-Damavandi,<sup>4</sup> Elizabeth C. Smyth,<sup>4</sup> Ruwaida Begum,<sup>4</sup> Paul Mitchell Dowsett,<sup>7</sup> Johann de Bono,<sup>8</sup> Paul V. Di Lorenzo,<sup>9</sup> Matteo Fassan,<sup>9</sup> Owen J. Sansom,<sup>10</sup> Suzanne Eccles,<sup>5</sup> Naureen Starling,<sup>4</sup> Chiara Braconi,<sup>4,9</sup> Andrea Sottoriva,<sup>2</sup> Simon P. Robinson,<sup>3</sup> David Cunningham

- 90% positive prediction
- 100% negative prediction

Ooft *et al.*, *Sci. Transl. Med.* **11**, eaay2574 (2019)

SCIENCE TRANSLATIONAL MEDICINE | RESEARCH ARTICLE

## CANCER

### Patient-derived organoids can predict response to chemotherapy in metastatic colorectal cancer patients

Salo N. Ooft<sup>1,2\*</sup>, Fleur Weeber<sup>1,2\*</sup>, Krijn K. Dijkstra<sup>1,2†</sup>, Chelsea M. McLean<sup>1,2†</sup>, Sovann Kaing<sup>1,2</sup>, Erik van Werkhoven<sup>3</sup>, Luuk Schipper<sup>1,2</sup>, Louisa Hoes<sup>1,2</sup>, Daniel J. Vis<sup>2,4</sup>, Joris van de Haar<sup>1,2,4</sup>, Warner Prevoo<sup>5</sup>, Petur Snaebjornsson<sup>6</sup>, Daphne van der Velden<sup>1,2†</sup>, Michelle Klein<sup>1,2</sup>, Myriam Chalabi<sup>1</sup>, Henk Boot<sup>7</sup>, Monique van Leerdam<sup>7</sup>, Haiko J. Bloemendaal<sup>8</sup>, Laurens V. Beerepoot<sup>9</sup>, Lodewyk Wessels<sup>2,4,10</sup>, Edwin Cuppen<sup>2,11,12</sup>, Hans Clevers<sup>2,13,14</sup>, Emile E. Voest<sup>1,2,7§</sup>



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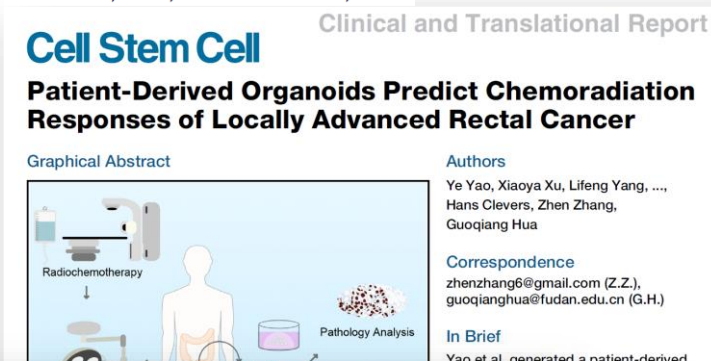
### Tumor Organoids Predict How Well Patients Respond to Cancer Drugs

Testing treatments on mini tumors may save time in identifying which therapies work best, a new study says

Feb 22, 2018  
RUTH WILLIAMS

In 100 percent of cases, if a drug didn't work on a patient's organoids, then it didn't work in the patient, and in nearly 90 percent of cases, if a drug did work on the organoids, then it worked in the patient.

Yao *et al.*, 2020, *Cell Stem Cell* **26**, 1–10



**Cell Stem Cell**  
Clinical and Translational Report

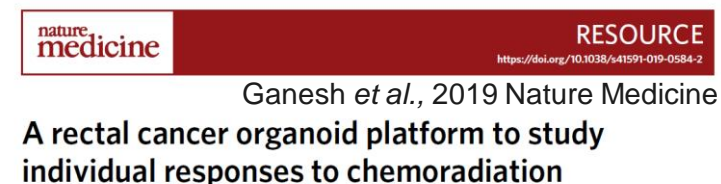
### Patient-Derived Organoids Predict Chemoradiation Responses of Locally Advanced Rectal Cancer

Graphical Abstract

Authors  
Ye Yao, Xiaoya Xu, Lifeng Yang, ..., Hans Clevers, Zhen Zhang, Guoqiang Hua

Correspondence  
zhenzhang6@gmail.com (Z.Z.), guoqianghua@fudan.edu.cn (G.H.)

In Brief  
Yao *et al.* generated a patient-derived organoid bank from patients with locally advanced rectal cancer and found that organoids had profiles that were similar to those of the patients. They found that PDOs can predict chemoradiation responses, which may have value as a companion diagnostic for LARC treatment.



**nature medicine**  
RESOURCE  
<https://doi.org/10.1038/s41591-019-0584-2>

### A rectal cancer organoid platform to study individual responses to chemoradiation

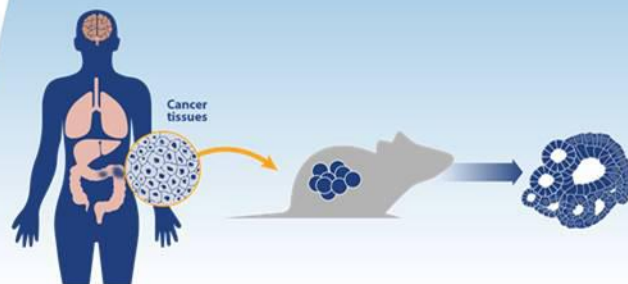
Ganesh *et al.*, 2019 *Nature Medicine*

Karuna Ganesh<sup>1,2,†</sup>, Chao Wu<sup>3,4,2†</sup>, Kevin P. O'Rourke<sup>5,6</sup>, Bryan C. Szeglin<sup>4,7</sup>, Youyun Zheng<sup>8,9</sup>, Charles-Etienne Gabriel Sauv  <sup>4</sup>, Mohammad Adileh<sup>4</sup>, Isaac Wasserman<sup>4</sup>, Michael R. Marco<sup>4</sup>, Amanda S. Kim<sup>10</sup>, Maha Shady<sup>8,9</sup>, Francisco Sanchez-Vega<sup>4,11</sup>, Wouter R. Karthaus<sup>3</sup>, Helen H. Won<sup>8,9</sup>, Seo-Hyun Choi<sup>4</sup>, Raphael Pelossof<sup>4</sup>, Afsar Barlas<sup>12</sup>, Peter Ntiamoh<sup>4</sup>, Emmanouil Pappou<sup>4</sup>, Arthur Elghouayel<sup>4</sup>, James S. Strong<sup>4</sup>, Chin-Tung Chen<sup>4</sup>, Jennifer W. Harris<sup>4</sup>, Martin R. Weiser<sup>4</sup>, Garrett M. Nash<sup>4</sup>, Jose G. Guillem<sup>4</sup>, Iris H. Wei<sup>4</sup>, Richard N. Kolesnick<sup>1</sup>, Harini Veeraraghavan<sup>13</sup>, Eduardo J. Ortiz<sup>14</sup>, Iva Petkovska<sup>14</sup>, Andrea Cercek<sup>2</sup>, Katia O. Manova-Todorova<sup>12</sup>, Leonard B. Saltz<sup>2</sup>, Jessica A. Lavery<sup>15</sup>, Ronald P. DeMatteo<sup>16</sup>, Joan Massagu  <sup>5</sup>, Philip B. Paty<sup>4</sup>, Rona Yaeger<sup>2</sup>, Xi Chen<sup>17</sup>, Sujata Patil<sup>18</sup>, Hans Clevers<sup>18</sup>, Michael F. Berger<sup>8,9</sup>, Scott W. Lowe<sup>5</sup>, Jinru Shia<sup>8,19</sup>, Paul B. Romesser<sup>10</sup>, Lukas E. Dow<sup>20</sup>, Julio Garcia-Aguilar<sup>4</sup>, Charles L. Sawyers<sup>3,21\*</sup> and J. Joshua Smith<sup>3,4,21\*</sup>



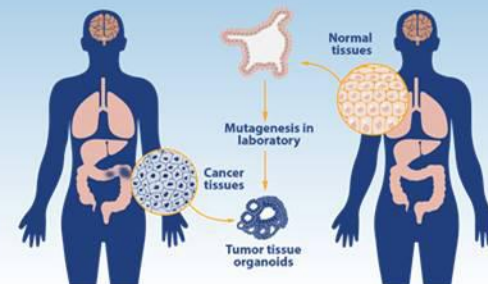
# CrownBio Organoid Biobank

**The only available organoid biobank for preclinical oncology developed using HUB's well-established, patented organoid technology**



## PDX-derived organoids (PDXO)

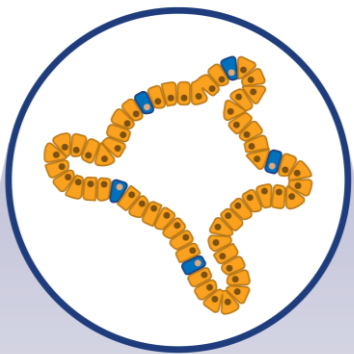
- Built using CrownBio PDX collection as a source of human tissue to complement HUB organoid collection
- **>180 models** in our master biobank over **15 cancer types**
- Matched *in vivo* PDX models
- HTS screening platform available
- Panels of models with oncogenic driver mutations



## HUB patient-derived organoids (PDO)

- Models transferred from HUB include breast, colorectal, lung, and pancreatic cancer organoids
- Rapidly expanding collection of **>180 models**
- Key collection features include:
  - Primary and metastatic matched pairs
  - Normal and tumor matched pairs
  - Models capturing heterogeneity of the patient tumor
  - Panels of models with oncogenic driver mutations

## Organoid



Derived from stem cells

Multiple cell lineages

Recapitulate organ  
physiological parameters

Long term culture

# Why use Use Tumor Organoids?

## Advantages

- Clinically predictive models
- Greater physiological relevance
- Scalable, amenable to biobanking & engineering
- Highly reproducible with automation achievable for HTS
- Normal and disease tissue organoids

## Limitations

- More complex to establish than 2D
- Automation has lower throughput than 2D
- Epithelial origins or model not yet available

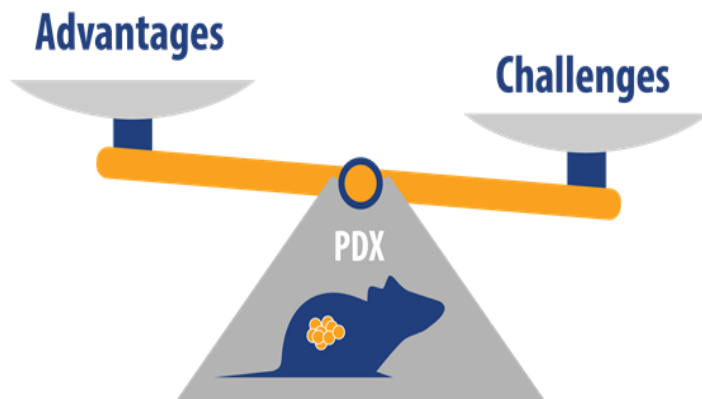
## Applications

- Matrix HTS or clinical trial in a dish
- Matched or labelled *in vivo* models
- Target validation, engineer TAA, biomarker identification
- Potential for matched normal tissue for on/off target effects
- Co-culture with immune cells or stromal cells for relevant TME

Advantages & Limitations

# **DO 3D SYSTEMS REPLACE IN VIVO MODELS?**

# Why Use *In Vivo* PDX Models?



## Advantages

- Clinically predictive models
- Greater physiological relevance and patient diversity
- Large biobanks enable mouse clinical trials to be conducted

## Limitations

- Ethical – clinical & animal welfare
- Longer to establish than organoids & lower throughput than 3D
- Favourable for more aggressive tumors
- Limited matched *in vitro* models
- Limited applicability to immuno-oncology

## Applications

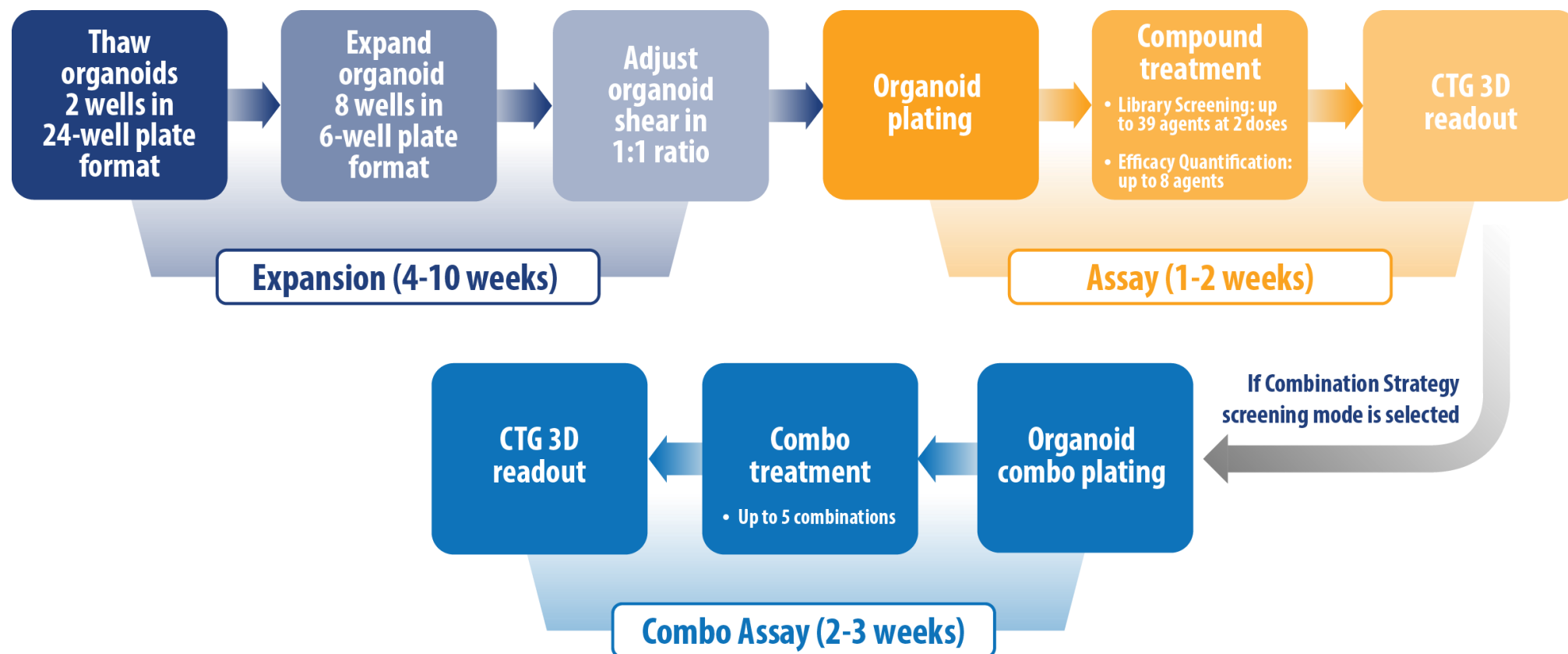
- Mouse clinical trial
- Efficacy or combination evaluation
- PK/PD
- Evaluate SoC, targeted agents, cellular therapies and more
- Humanized models to evaluate immune modulators

Feasibility Study

# **EVALUATE SYNERGISTIC EFFECTS USING A PDXO SCREENING PLATFORM**

# Organoid Screen Workflow

- Screen multiple compounds using either a single concentration, multiple concentrations or combinations across multiple patients



# Study Objective

- Evaluate the feasibility of a **PDXO-based screening platform** to test potential synergistic effects of 2 compounds
- Validate **matrix screen** set up
  - Multiple combination strategies, dosing and models tested in parallel
- Analyze the data using **two independent mathematical models** for additional confidence





# Background and Significance

- Synergies between the WEE1 inhibitor MK-1775 and the CHK inhibitor MK-8776, and each with gemcitabine, have been reported for the treatment of **p53 mutant pancreatic cancer** as well as other cancer types

## Mechanistic Distinctions between CHK1 and WEE1 Inhibition Guide the Scheduling of Triple Therapy with Gemcitabine

Siang-Boon Koh, Yann Wallez, Charles R. Dunlop, Sandra Bernaldo de Quirós Fernández, Tashinga E. Bapiro, Frances

DOI: 10.1158/0008-5472.CAN-17-3932 Published June 2018 

## MK-1775, a Potent Wee1 Inhibitor, Synergizes with Gemcitabine to Achieve Tumor Regressions, Selectively in p53-Deficient Pancreatic Cancer Xenografts

N.V. Rajeshkumar, Elizabeth De Oliveira, Niki Ottenhof, James Watters, David Brooks, Tim Demuth, Stuart D. Shumway, Shinji Mizuarai, Hiroshi Hirai, Anirban Maitra, and Manuel Hidalgo

DOI: 10.1158/1078-0432.CCR-10-2580 Published May 2011

## Dose Escalation Trial of the Wee1 Inhibitor Adavosertib (AZD1775) in Combination With Gemcitabine and Radiation for Patients With Locally Advanced Pancreatic Cancer

[Kyle C. Cuneo](#), MD<sup>1</sup>; [Meredith A. Morgan](#), PhD<sup>1</sup>; [Vaibhav Sahai](#), MBBS, MS<sup>1</sup>; [Matthew J. Schipper](#), PhD<sup>1</sup>; [Leslie A. Parsels](#), PhD<sup>1</sup>; [Joshua D. Parsels](#)<sup>1</sup>; ...

## Unique functions of CHK1 and WEE1 underlie synergistic anti-tumor activity upon pharmacologic inhibition

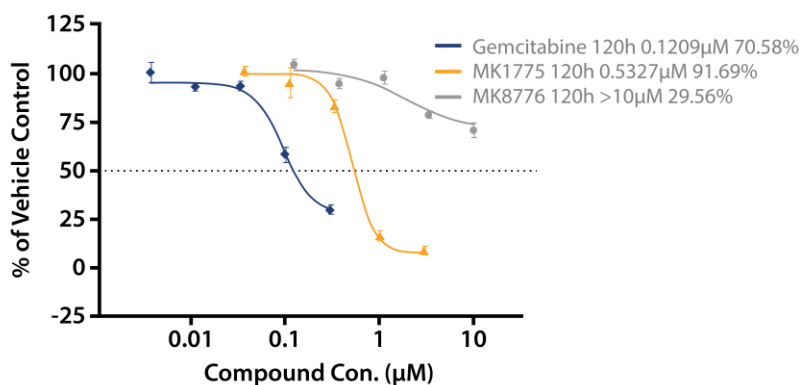
[Amy D Guertin](#), [Melissa M Martin](#), [Brian Roberts](#), [Melissa Hurd](#), [Xianlu Qu](#), [Nathan R Miselis](#), [Yaping Liu](#), [Jing Li](#), [Igor Feldman](#), [Yair Benita](#), [Andrew Bloecher](#), [Carlo Toniatti](#) & [Stuart D Shumway](#) 

*Cancer Cell International* 12, Article number: 45 (2012) | [Cite this article](#)

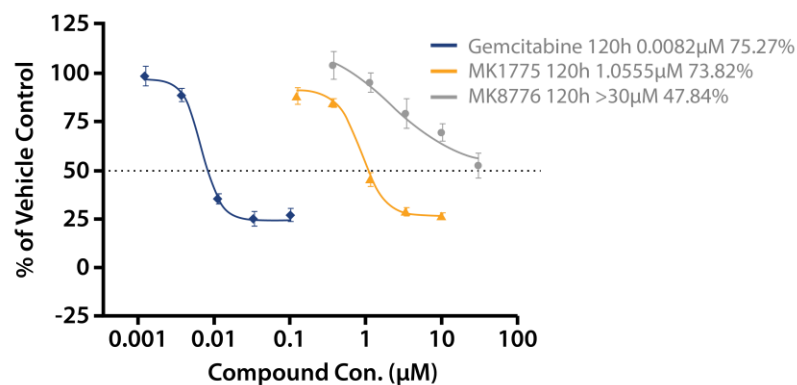
8270 Accesses | 51 Citations | 1 Altmetric | [Metrics](#)

- *p53* mutant PDXO models selected similar models to published studies
  - PA5389B and PA1252B with a *p53*<sup>R282W</sup> and *p53*<sup>R273H</sup> missense mutation, respectively
  - PA2847B with a *p53*<sup>W91Ter</sup> nonsense mutation
- Dose response curves initially established for single agents
- All models were:
  - Sensitive to gemcitabine [ $IC_{50} < 0.2 \mu M$ ] and MK-1775 [ $IC_{50} < 1 \mu M$ ]
  - Poor sensitivity to MK-8776 [ $IC_{50} > 10 \mu M$ ]

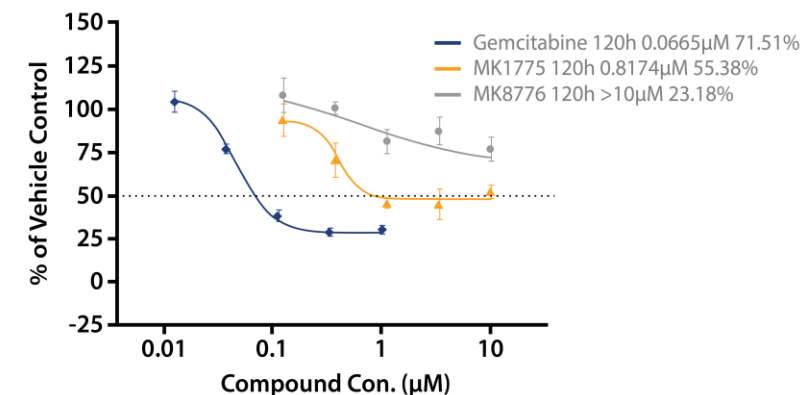
**PA1252B Dose-Response Curve**



**PA5389B Dose-Response Curve**



**PA2847B Dose-Response Curve**



# Combination Study Design

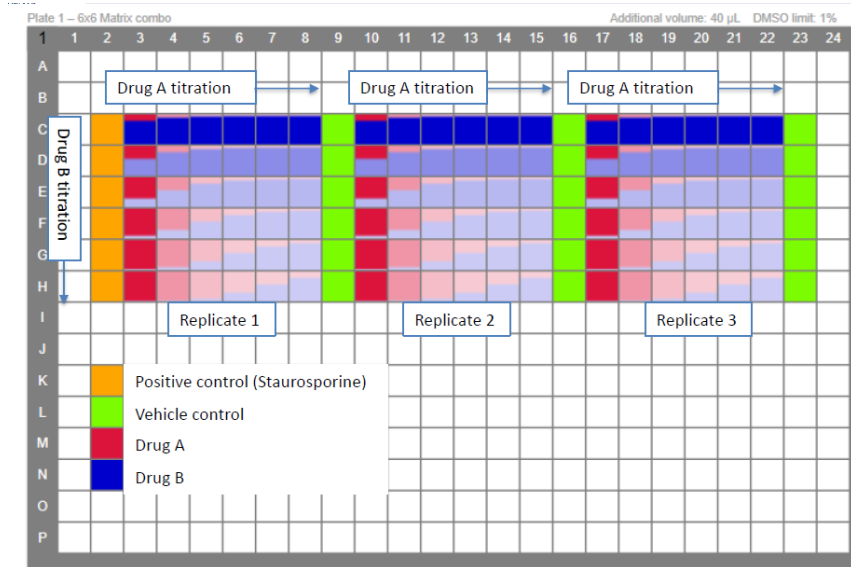
- 3 combination strategies
- 6-point titration of each inhibitor combined in a 6×6 matrix format, for 5 days
- 3 different PDXO models
- Cell viability measured
- 2 independent mathematical models of synergy (Bliss and Loewe) used to assess the combination effect
  - Synergy score >5 indicates synergistic effect
  - Synergy score <5 indicates antagonism

	Drug 1	Drug 2
<b>Combo 1</b>	MK-1775	MK-8776
<b>Combo 2</b>	Gemcitabine	MK-8776
<b>Combo 3</b>	Gemcitabine	MK-1775

6×6 matrix, 1 model/plate

MK-1775: max [Conc] = 10μM; dilution factor 3

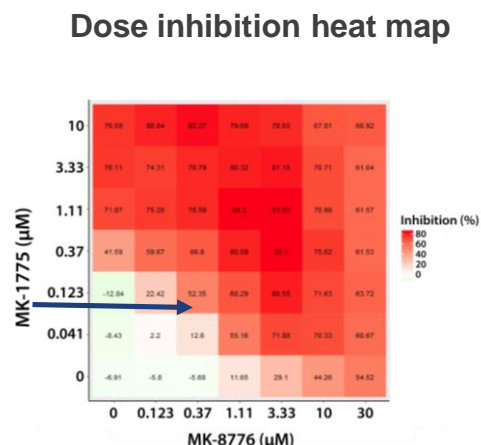
MK-8776: max [Conc] = 30μM; dilution factor 3



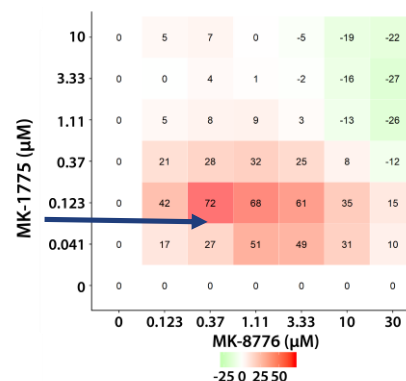
# Example of Matrix Design Results

- Combination of MK-1775+MK-8776 analyzed by both Bliss and Loewe methods showed strong synergy at low concentrations (PA5389B pancreatic cancer organoid)

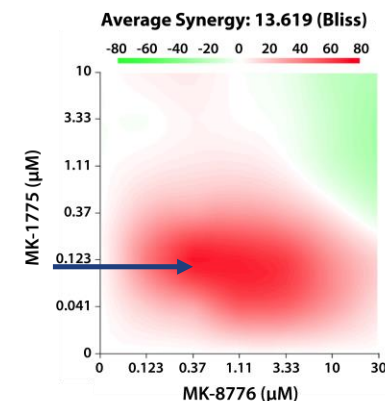
Bliss



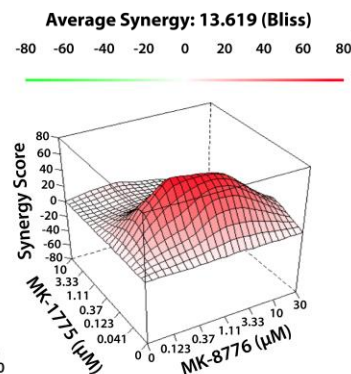
**Synergy score heat map**



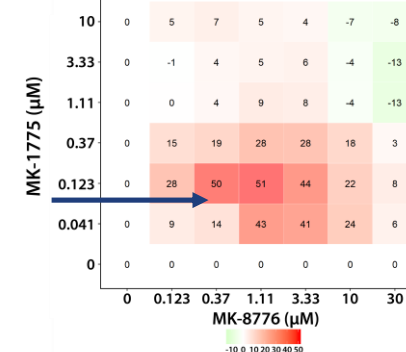
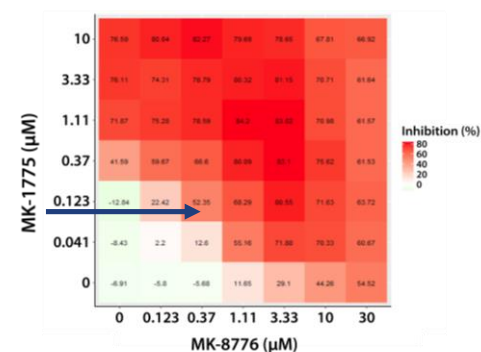
**2D contour map**



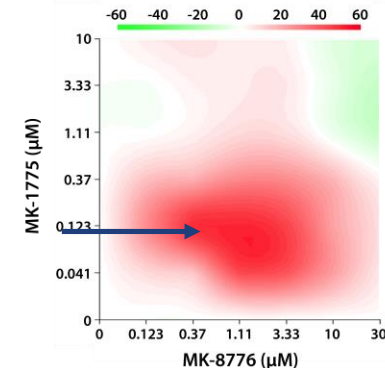
**3D response surface plot**



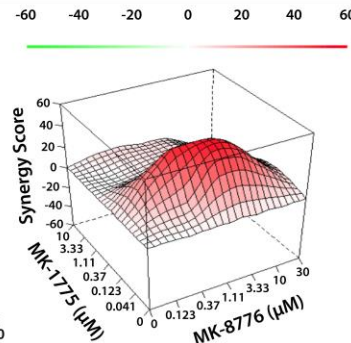
Loewe



**Average Synergy: 12.822 (Loewe)**



**Average Synergy: 12.822 (Loewe)**



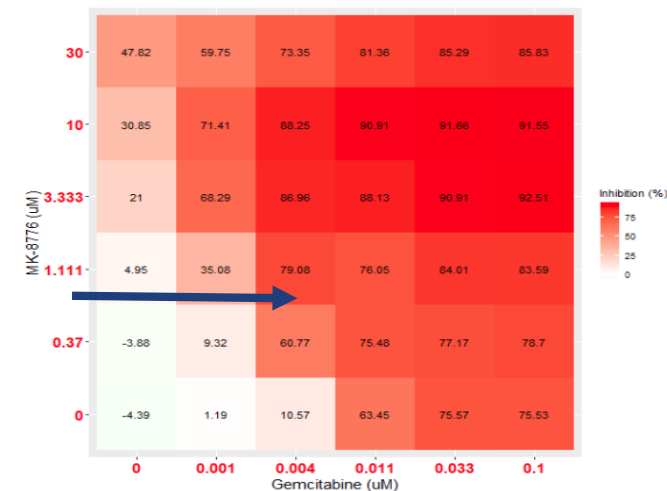
## Combination with Gemcitabine

- In combination with SoC gemcitabine, the highest synergy scores (>30) were seen with MK-8776/gemcitabine combo, with both mathematical models
- However, some low average values were also observed, suggesting antagonism with certain dose combinations
- MK-8776 achieved a 67-88% growth inhibition at the non-inhibitory concentration of 1.111 $\mu$ M, when in combination with 11-33nM gemcitabine

Model	MK-1775/Gemcitabine				MK-8776/Gemcitabine			
	Highest Synergy Score		Average Synergy Score		Highest Synergy Score		Average Synergy Score	
	Bliss	Loewe	Bliss	Loewe	Bliss	Loewe	Bliss	Loewe
PA5389B	48.8	44.57	8.40	14.95	64.08	56.78	19.92	21.49
PA2847B	32.19	15.34	-4	-5.04	60.1	42.64	11.55	-4.80
PA1252B	58.72	44.75	14.23	12.06	79.31	32.27	70.89	27.61

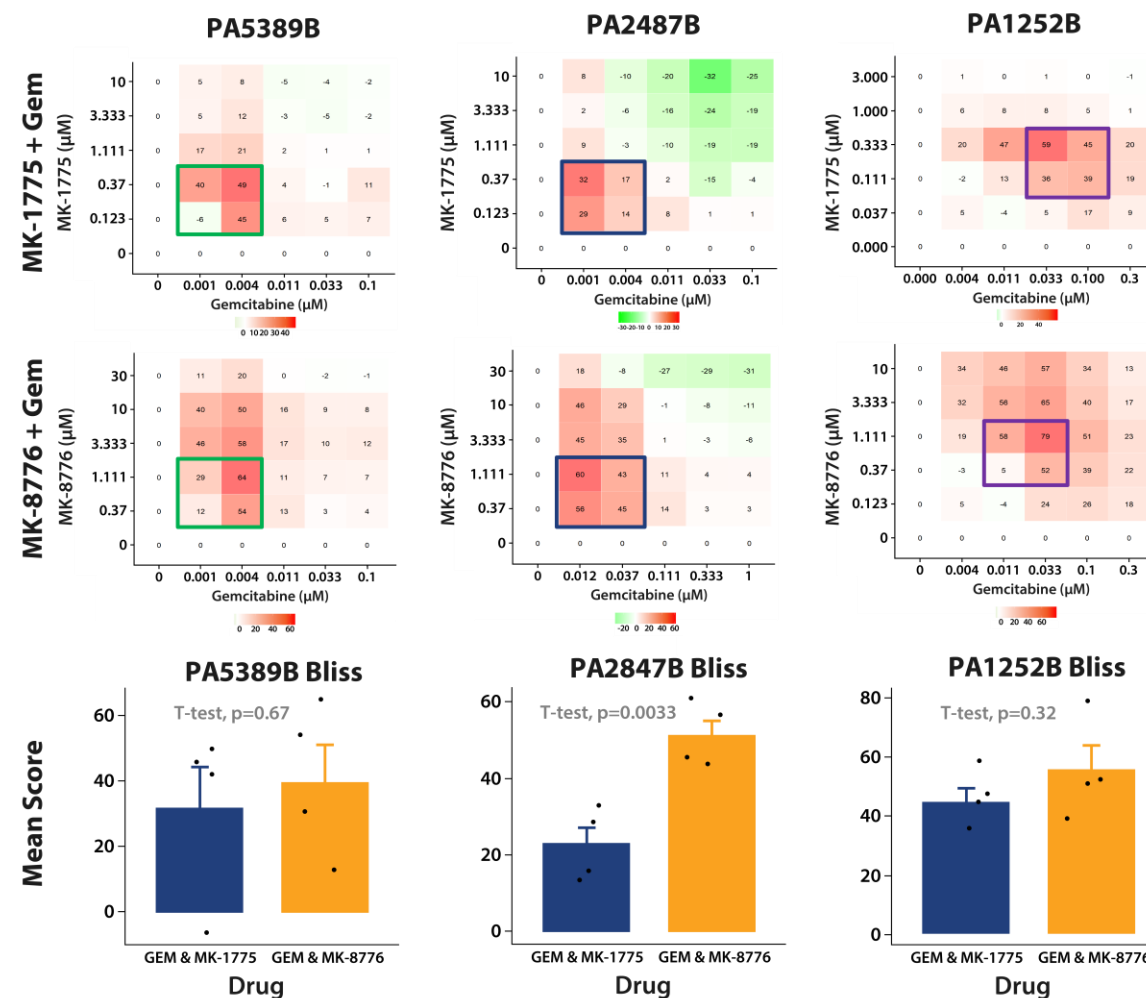
Synergy score >5 indicates synergist effects  
Synergy score <5 indicates antagonistic effects

PA5389B dose inhibition heat map



# Comparison of Synergy

- Quadrant with highest synergy scores was found to be at lower drug concentrations for the models with p53 missense mutation (PA5389B and PA2487B), compared to PA1252B
- Overall a higher synergy score was observed in the p53 mutated models at lower concentrations
- Bliss analysis also showed MK-1775 + gemcitabine to have a significantly higher mean synergy score than MK-8776 in PA2487 organoid model

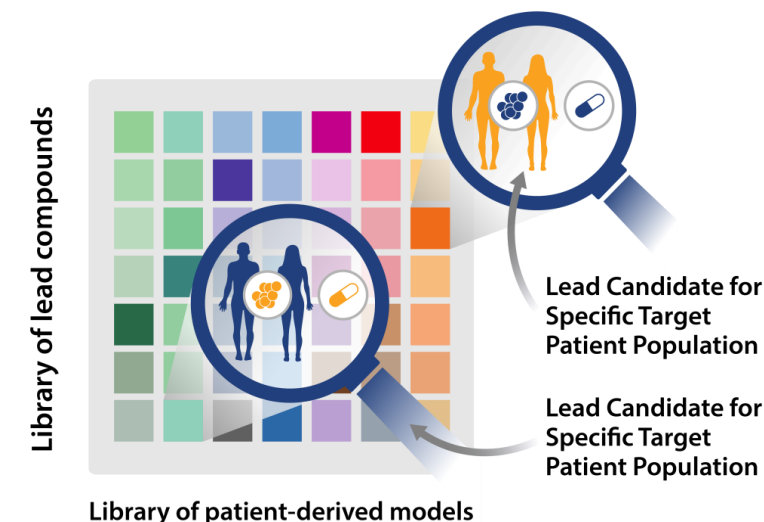




# Study Conclusions

- Synergistic effects of MK-1775 (WEE1 inhibitor) and MK-8776 (CHK inhibitor) were assessed in highly patient-relevant p53 mutant pancreatic cancer PDXO models
- Results indicated:
  - All pancreatic cancer PDXO models were sensitive to gemcitabine and MK-1775, and poorly sensitive to MK-8776
  - The combination of MK-1775 and MK-8776 shows very strong synergistic effect in all models tested
  - The synergy observed between MK-8776/gemcitabine was significantly greater than MK-1775/gemcitabine in one of the p53 mutated models (PA2847) at low doses, however antagonism was observed at higher concentrations
- The rapid screening of patient-relevant organoids in a combination matrix approach enables synergistic profiling to be easily conducted, providing valuable insight into combination strategies

- Patient-relevant models can bridge the gap between the lab and the clinic
- Adoption during early *in vitro* stages can be achieved with 3D models
- However, some 3D tumor spheroid models can lack reproducibility, efficiency and translatability
- In comparison, 3D organoids are “mini-organs in a dish” derived from tissue stem cells, that are highly predictive of the *in vivo* and clinical response
- Adoption of organoids could potentially improve the drug discovery workflow, requiring expansion of biobanks and matched models
- Predicting *in vivo* responses will refine the application of *in vivo* models



- Tumor Organoid Applications to Drug Discovery and Development Resource Library

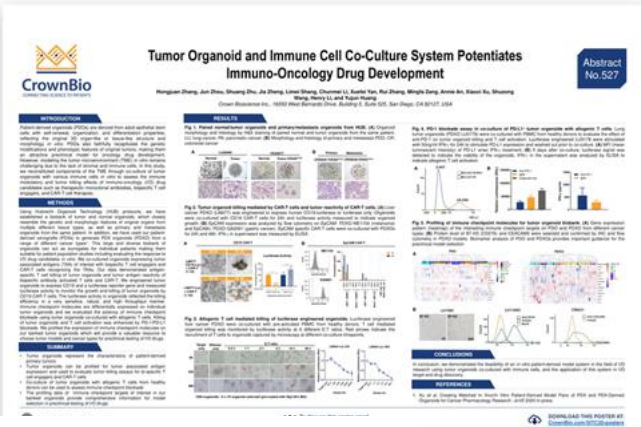


**New Tumor Organoid Immuno-Oncology Platform**

organoids

immune cells

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**WHITE PAPER**

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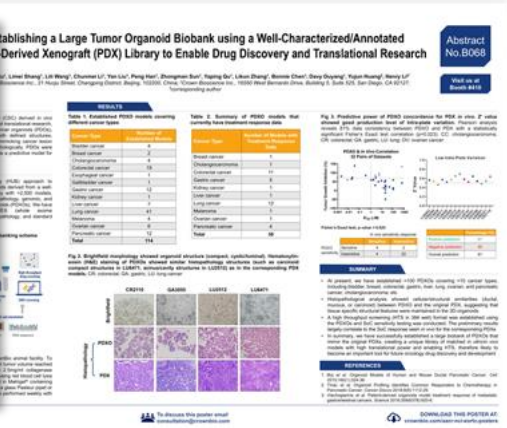


**Meet the Team**

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Lead Scientist, CBNL

**Dr. Xiaoxi Xu**  
Head of Organoid Platform, CBNL

**Dr. Yujun Huang**  
Head of Immuno-Oncology Platform, CBSD



**Establishing a Large Tumor Organoid Biobank using a Well-Characterized/Annotated Cell-Derived Xenograft (PDX) Library to Enable Drug Discovery and Translational Research**

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